

Accelerating growth of *Pseudomonas putida* through media optimization to improve rhamnolipid production

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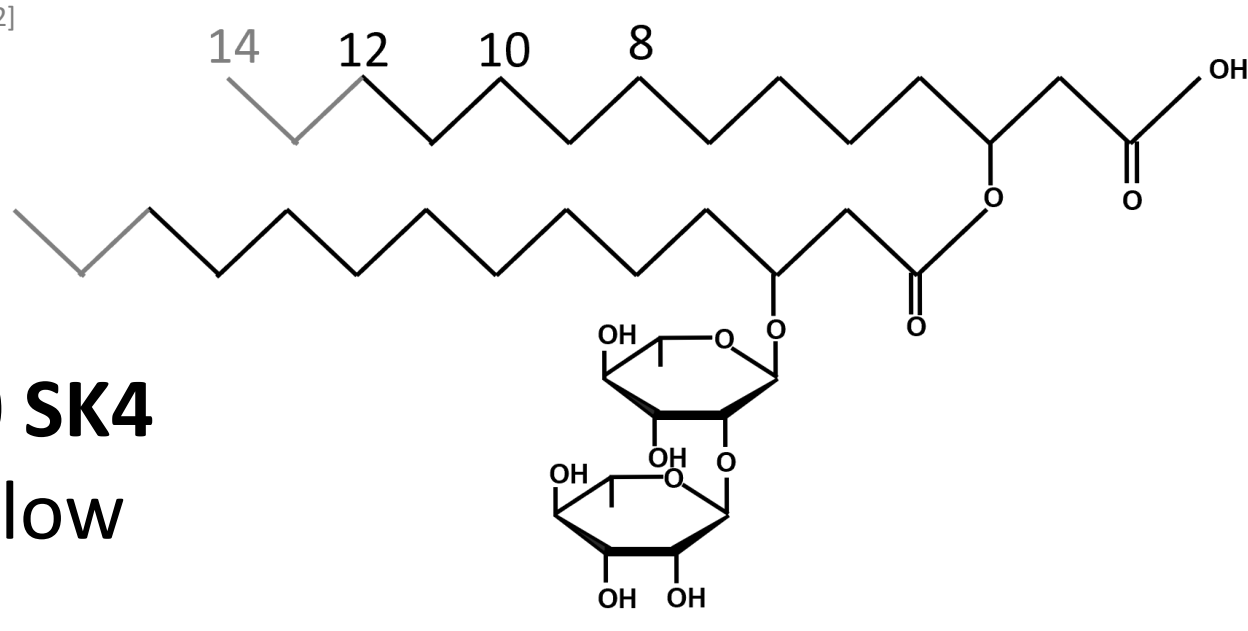
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Rhamnolipids

- Amphiphilic molecules
- Possible **applications**: plant protection, bioremediation [1], [2]
- Naturally produced by e.g. *Pseudomonas aeruginosa* [3]
- Recombinant production in *Pseudomonas putida* KT2440 SK4
- Current media developed for low cell densities [4]



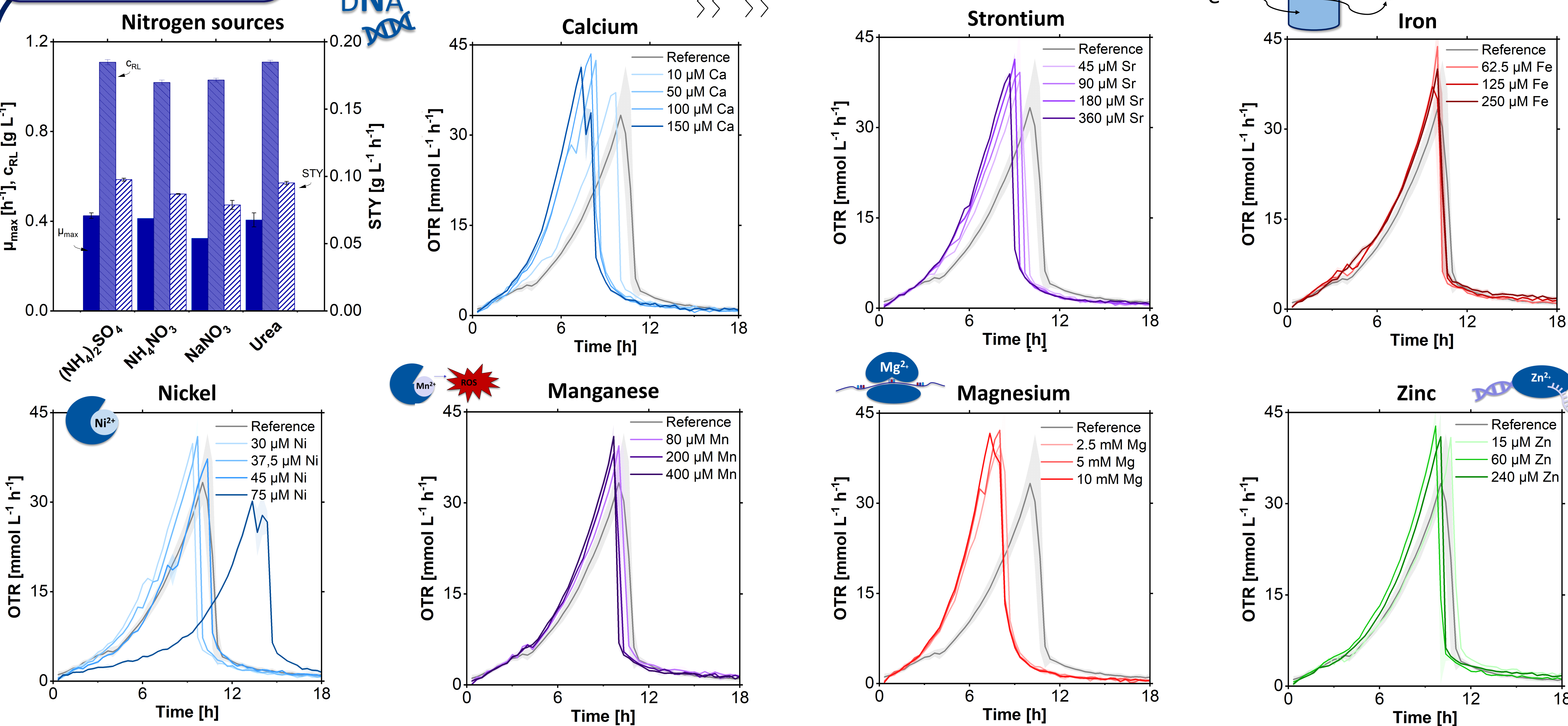
Aim

- Investigation of **influencing factors** on rhamnolipid production
- Nitrogen source
- Trace elements
- Development of an optimized medium for **high-cell-density** cultivations

No improve

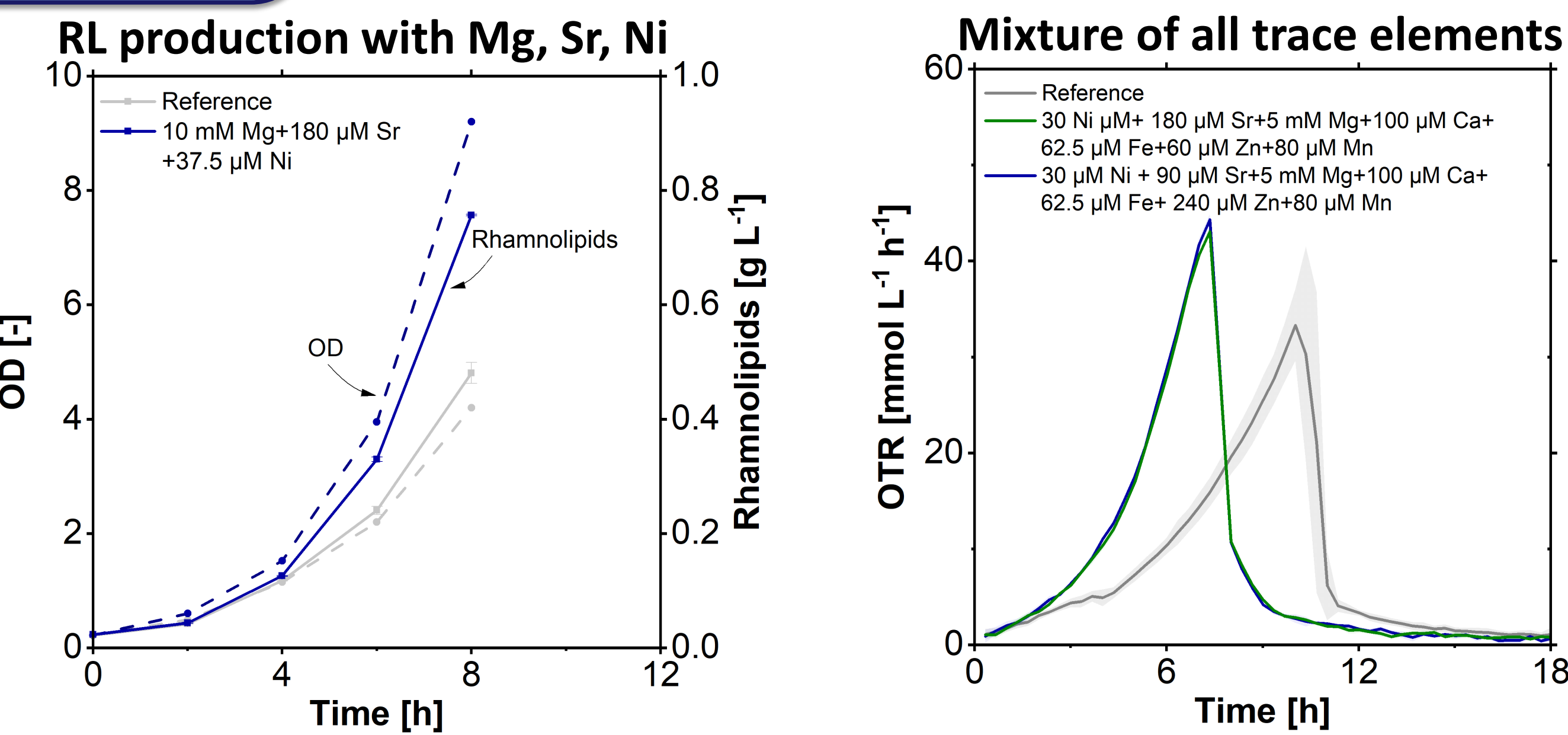
- Tungsten
- Aluminium
- Cobalt
- Copper
- Molybdenum
- Selenium
- Boron

Influencing factors



Cultivation conditions: Cultivation of *P. putida* KT2440 SK4 with different nitrogen sources and trace elements. Test of different nitrogen sources ($(NH_4)_2SO_4$, NH_4NO_3 , $NaNO_3$ and Urea) in a shake flask experiment in MSM-medium with 10 g L⁻¹ glucose, N=250 rpm, T = 30 °C, OD = 0.2, pH_{start} = 7.0. Test of different trace elements (Ca, Sr, Fe, Ni, Mn, Mg, Zn) with varying concentrations in a 96-well microTOM (Kühner Shaker GmbH, Biersfelden, Switzerland) experiment in MSM-medium with 10 g L⁻¹ glucose + 5 g L⁻¹ $(NH_4)_2SO_4$, N = 300 rpm, T = 30 °C, OD = 0.2, pH_{start} = 7.0.

Mixtures

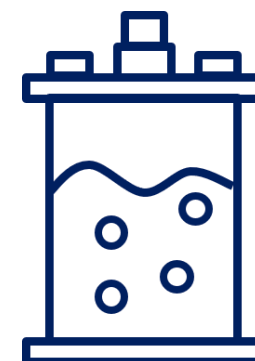


Faster growth and rhamnolipid production with mixture of Mg, Sr and Ni ($\mu_{max} = 0.48 \pm 0.00$ h⁻¹, $STY = 0.09 \pm 0.00$ g L⁻¹ h⁻¹) observed compared to reference ($\mu_{max} = 0.41 \pm 0.01$ h⁻¹, $STY = 0.04 \pm 0.00$ g L⁻¹ h⁻¹). Final mixtures of all trace elements showed **1,6-fold faster growth**.

Cultivation conditions: Shake flask and microTOM cultivation of *P. putida* KT2440 SK4 with varying TE concentration. Same cultivation conditions used as in the experiments described above.

Conclusion & Outlook

- $(NH_4)_2SO_4$ best performing nitrogen source
- Faster growth through addition of **Mg, Sr, Ca, Mn, Ni, Zn, Fe**
- Mixture showed faster growth and rhamnolipid production
- Next steps:
 - Detailed examination of final mixture necessary
 - **High-cell-density** cultivations



References

- [1] Kiran et al. (2016), Crit Rev Biotechnol, 3: p.399-415
- [2] Bredenbruch et al. (2024), Pestic Biochem Physiol, 204
- [3] Jarvis and Johnson (1949), J Am Chem Soc 71:4124-4126
- [4] Hartmans et al. (1989), Appl. Environ. Microbiol., 55: 2850-2855